

REVIEW

Adiponectin, Leptin and Cardiovascular Disorders

Shangang Zhao, Christine M. Kusminski, Philipp E. Scherer 

ABSTRACT: The landmark discoveries of leptin and adiponectin firmly established adipose tissue as a sophisticated and highly active endocrine organ, opening a new era of investigating adipose-mediated tissue crosstalk. Both obesity-associated hyperleptinemia and hypoadiponectinemia are important biomarkers to predict cardiovascular outcomes, suggesting a crucial role for adiponectin and leptin in obesity-associated cardiovascular disorders. Normal physiological levels of adiponectin and leptin are indeed essential to maintain proper cardiovascular function. Insufficient adiponectin and leptin signaling results in cardiovascular dysfunction. However, a paradox of high levels of both leptin and adiponectin is emerging in the pathogenesis of cardiovascular disorders. Here, we (1) summarize the recent progress in the field of adiponectin and leptin and its association with cardiovascular disorders, (2) further discuss the underlying mechanisms for this new paradox of leptin and adiponectin action, and (3) explore the possible application of partial leptin reduction, in addition to increasing the adiponectin/leptin ratio as a means to prevent or reverse cardiovascular disorders.

Key Words: adiponectin ■ cardiovascular diseases ■ hypoadiponectinemia ■ leptin ■ obesity

OBESITY AND CARDIOVASCULAR DYSFUNCTION

Currently, the increasing rate of obesity rises steeply and now reaches global pandemic proportions.¹ Obesity is one of the major health issues and is a crucial contributor to the global burden of chronic disease and disability. Obesity positively correlates with various metabolic challenges, including insulin resistance, type 2 diabetes, non-alcoholic fatty liver disease, cardiovascular dysfunction, and certain types of cancers.² The health consequences of such metabolic dysfunctions range from reduced quality of life to premature death. Of particular concern is the increased incidence of cardiovascular dysfunction, a group of diseases of the heart and blood vessels, mainly including coronary heart disease, cerebrovascular disease, peripheral arterial disease, rheumatic heart disease, congenital heart disease, deep vein thrombosis, and pulmonary embolism; it is the leading cause of death for individuals of most racial and ethnic groups in the United States.³ Early-childhood obesity by 11 to 12 years, is positively associated with the development of cardiovascular dysfunction, highlighting the crucial link to obesity.⁴ The identification of the key causative factors

that mediate obesity-associated cardiovascular disorders is an urgent and still unmet need, even though this has been extensively explored over the past 2 decades.⁵ Much interest has been directed towards a better understanding of the pathological expansion of fat mass, which may represent a crucial link between obesity and cardiovascular disease.⁶

THE INHERENT HETEROGENEITY OF ADIPOSE TISSUE

Adipose tissue shows a high degree of heterogeneity.⁷ This is reflected by many aspects, such as the different adipose tissue depots located throughout the body, their unique individual cellular characteristics of the adipocytes and their diverse cellular composition.⁸ Simply based on its location, adipose tissue can be subdivided into numerous categories. These include depots in subcutaneous, visceral, supraclavicular, anterior cervical, axillary, anterior subcutaneous, suprascapular, supraspinal, ventral spinal, infrascapular, dermal, pericardial and perirenal regions.^{9,10} Of note, due to their close proximity to organs critically involved in the pathophysiology of the cardiovascular

Correspondence to: Philipp E. Scherer, PhD, Touchstone Diabetes Center, Department of Internal Medicine and Cell Biology, The University of Texas Southwestern Medical Center, 5323 Harry Hines Boulevard, Dallas, Texas 75390-8549. Email philipp.scherer@utsouthwestern.edu

For Sources of Funding and Disclosures, see page 145.

© 2020 American Heart Association, Inc.

Circulation Research is available at www.ahajournals.org/journal/res

Nonstandard Abbreviations and Acronyms

AdipoR	adiponectin receptor
AdipoRon	an AdipoR1 and AdipoR2 agonist
CB1	cannabinoid receptor 1
CRP	C-Reactive Protein
db/db mice	mice deficient in leptin receptor
HMW	high molecular weight
LepRb	Leptin receptor isoform b
ob/ob mice	mice deficient in leptin
PPAR	peroxisome proliferator-activated receptor
ROS	reactive oxygen species
SGLT-2	sodium-glucose-linked transporter-2
STAT3	signal transducer and activator of transcription 3

system, the pericardial depot is closely associated with cardiovascular function.¹¹ Moreover, in rodents, fat-depots display a high degree of topological similarity to the depots in humans.¹⁰ This adds much credibility to the use of murine model systems to study the progression of obesity in humans. Nevertheless, despite sharing a similar location, rodent and human retroperitoneal fat depots show some differences. In humans, the retroperitoneal fat often encapsulates the kidney, adheres tightly to the renal capsule and invades the renal sinuses; while in the mouse, this fat depot simply surrounds the kidney.¹² Beyond the differences in location, adipose tissue also has a complex cellular composition. The various depots consist of mature adipocytes, mesenchymal progenitor/stem cells, preadipocytes, endothelial cells, mural cells, T cells and macrophages.^{13,14} Different cellular populations within adipose tissue have very distinct functions. For example, preadipocytes can be differentiated into mature adipocytes by providing a classical differentiation cocktail. It is this population of cells that are the major source for generating new mature adipocytes.¹⁵ Endothelial cells share the same progenitor cells with mature adipocytes, and endothelial cells can be rapidly incorporated into vessels; for instance, to promote both postischemic neovascularization in nude mice and vessel-like structure formation in Matrigel plugs.¹⁶ Endothelial cells also play an important role in the development of insulin resistance.¹⁷ As such, a better understanding of adipose tissue heterogeneity will certainly help identify the critical players in mediating adipose tissue-associated cardiovascular disorders.

Adipose tissue has the unique ability to expand to an almost unlimited extent, despite not being transformed.¹⁸ In response to excess energy supply, adipose tissue undergoes complete remodeling. This involves activation of a highly coordinated process among several cell types, including mural cells, macrophages and preadipocytes.¹⁹

How this remodeling occurs during expansion is the key difference between healthy adipose tissue expansion versus unhealthy expansion. Failure to adequately remodel while expanding results in chronic, unresolved inflammation and metabolic dysfunction.^{19–21} The expansion of adipose tissue can occur via 2 distinct mechanisms: (1) the increase in adipocyte size (hypertrophy), or (2) the generation of more adipocytes through precursor cell differentiation, a process termed adipogenesis (hyperplasia).²² Hypertrophy, with larger adipocytes, is associated with a poor metabolic response, which includes unleashed lipolysis, altered patterns of adipokine secretion and enhanced proinflammatory cytokine secretion.²³ In contrast, hyperplasia is a much healthier form of adipose tissue remodeling, which generates many new, smaller adipocytes through recruitment and differentiation of preadipocytes.²⁴ In response to chronic metabolic challenge, such as high-fat diet feeding, both forms of adipose tissue remodeling can effectively be monitored and tracked in various fat-depots, by using the Adipo-chaser mouse model system.²⁵ This allows us, for the first time, to clearly differentiate between preexisting and new adipocytes in response to any kind of metabolic challenge.

The quantity and quality of adipose tissue are equally important in light of the obesity-associated increased risk for many accompanying health issues.²⁶ Too little adipose tissue, resulting from chronic weight loss or genetic issues in fat development, leads to severe congenital or acquired lipoatrophy.²⁷ The latter is a classical disorder that is characterized by insulin resistance, hypertriglyceridemia, nonalcoholic fatty liver disease, and cardiovascular disorders. In light of the continuum between lipoatrophy and obesity, maintaining the proper amount of adipose tissue mass is clearly crucial in preventing multiple metabolic sequelae of dysfunctional adipose tissue, such as cardiovascular disorders. Even with comparable amounts of adipose tissue, obese individuals display a range of metabolic phenotypes. The majority of obese individuals develop insulin resistance, hypertriglyceridemia, liver steatosis, hypertension, and cardiovascular disorders. However, a subset of obese individuals maintain a high degree of insulin sensitivity, thus rendering protection from various metabolic disorders. This favors the idea of metabolically healthy obesity, in contrast to metabolically abnormal obesity.^{28,29} This idea is well established in clinical observations and strongly supported by rodent studies.^{30–33} Much remains to be done to delineate the exact mechanisms that determine these dramatic metabolic differences. Over the decades, numerous studies have demonstrated that adipose tissue fibrosis and adipokine production are key determinants of pathological metabolic sequelae. This indicates that not only quantity, but also the quality of adipose tissue is equally important in exerting its effects.³⁴

Historically, adipose tissue was predominantly viewed as a relatively inert energy reservoir. In fact, having

vast energy stores is an advantage for survival during reduced caloric availability. However, in more recent times of fuel surplus, the role of adipose tissue as an energy store was less of a research focus. As such, for many decades, the overall interest of adipose tissue was somewhat overlooked. The discoveries in the 1990s of leptin and adiponectin revitalized and helped reshape adipose tissue from simply being an energy reservoir, to a sophisticated and highly active endocrine organ. This opened a new era of exploring adipose tissue-mediated crosstalk with other organs.^{35,36} In addition to leptin and adiponectin, adipose tissue secretes a large panel of other adipokines, cytokines, metabolites, and exosomes. Together, these form a unique secretome response that mediates inter-organ communication. This concept has been covered in our recent review.³⁷ Here, we will focus on leptin and adiponectin and their roles in cardiovascular function.

LEPTIN

Leptin was discovered in 1994³⁸ as a 16 kDa nonglycosylated protein that is predominately secreted from adipose tissue.³⁸ Other tissues, including skeletal muscle,³⁹ gastric mucosa,⁴⁰ placenta,⁴¹ heart, mammary, and salivary glands⁴² can produce small amounts of leptin under certain conditions. However, adipose tissue seems to be the predominant source for circulating leptin. This is due to the fact that deletion of the *Lep* gene exclusively in adipose tissue, leads to undetectable levels of the protein in circulation.⁴³ The circulating levels of leptin are highly proportional to the amount of adipose tissue mass, that is, the higher the fat mass, the higher the circulating levels of leptin.⁴⁴ For instance, female subjects, with overall larger relative fat mass, tend to exhibit 2-fold higher leptin levels in circulation, when compared with males of similar body weight.³⁸

As a pleiotropic hormone,⁴⁵ leptin regulates many physiological processes, including food intake, non-shivering thermogenesis,^{46,47} reproduction, hemostasis, angiogenesis, arterial pressure control,⁴⁸ and neuroendocrine and immune function.⁴⁹ To fulfill its physiological role, leptin must bind to the LepRb (long isoform of its receptor), which is highly enriched in the hypothalamic region of the brain and to a lesser extent in peripheral tissues and macrophages.^{50,51} To date, it is still unclear what the precise leptin-sensing mechanism is, that is, how and when the central nervous system communicates to the peripheral tissues in response to leptin, and vice versa. Two prevailing models have been proposed.⁵² In the first model, the levels of adipocyte-derived leptin in circulation are proportionally increased with the increase in fat mass. It is both these factors that activate a response from the central nervous system, which ultimately prompts a corresponding increase in energy expenditure, concomitant with a reduction in food

intake.⁵² Firm evidence of this model lies in the fact that elevating the levels of leptin in young mice, in a dose-dependent manner, reduces food intake and increases core body temperature.⁵³ In contrast, the second model builds upon the notion that the signal sensed, is actually a drop, rather than an increase in leptin. This drop in leptin levels prompts a physiological response in energy regulation and reproduction.⁵⁴ The latter model is best demonstrated by starvation-induced leptin reduction and its associated physiological response, which can be largely reversed by exogenous leptin supplementation. Both models are well supported by numerous experimental observations. As such, it is difficult to argue which model is superior to the other. Given that the common factor in the 2 models is a change in circulating leptin levels, we believe that leptin oscillations, within their physiological range, are the critical determinant of leptin's overall function. On a daily basis, the circulating levels of leptin are relatively stable. As such, this argues against a physiological role for leptin in regulating acute daily food intake.⁵⁵ Rather, in subjects of normal body weight, a high-fat meal can typically trigger postprandial changes in the circulating levels of leptin. Conversely, in obese individuals, this postprandial response in leptin levels is significantly flattened and delayed.⁵⁶

LEPTIN'S PARADOXICAL EFFECTS ON CARDIOVASCULAR DISORDERS

Given the robust effects of leptin on food intake and energy expenditure, leptin therapy (by exogenously increasing circulating leptin levels) once promised to be a cure for diet-induced obesity and its associated metabolic disorders.⁵⁷ However, obesity-associated hyperleptinemia and leptin resistance rendered leptin therapy largely ineffective for the treatment of diet-induced obesity.⁵⁸ In the context of cardiovascular function, leptin exerts both dichotomous and paradoxical effects (Figure 1). In most cases, hyperleptinemia is positively correlated with unfavorable outcomes in cardiovascular disorders.^{59,60} However, under some circumstances, leptin can elicit cardioprotective effects, by reducing cardiomyocyte hypertrophy and apoptosis.⁶¹

A fully intact leptin system exists in all regions of the heart; this includes leptin synthesis and a fully functional long form of its receptor.⁶² As such, leptin signaling is necessary and indispensable for maintaining normal cardiac function. In young healthy men, a beneficial inverse correlation between measures of carotid wall thickness and circulating leptin is evident; thereby supporting a vascular protective role for leptin.⁶³ A deficiency in leptin itself (ie, *ob/ob* mice), or its receptor (ie, *db/db* mice) results in massive obesity and severe cardiovascular dysfunction.⁶⁴ The addition of leptin to *ob/ob* mice essentially restores normal thickness of the left ventricle; with this effect being independent of body weight.⁶⁵ Furthermore,

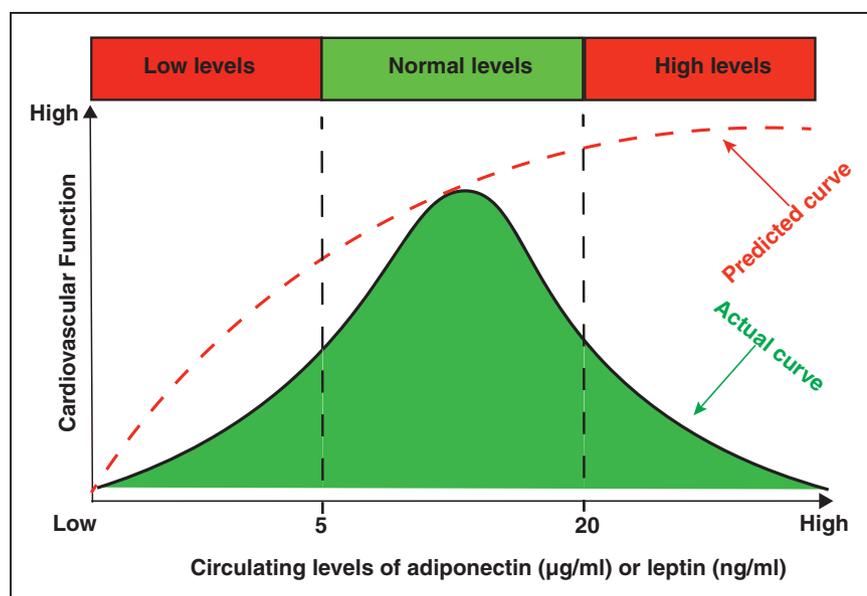


Figure 1. The paradoxical effects of adiponectin and leptin in regulating cardiovascular function.

Low levels of adiponectin and leptin are positively associated with severe cardiovascular disorders. Thus, it is predicted that high adiponectin and high leptin levels, beyond physiological (normal) levels, could greatly improve cardiovascular function, as shown by the red dashed curve. However, in most cases, high circulating leptin and adiponectin levels do not show any beneficial effects. Rather, both can be detrimental for cardiovascular function, similar to the conditions with low circulating levels, as shown in the green area, referred to as paradoxical effects.

restoring leptin receptor expression exclusively in cardiac tissue in *db/db* mice, reduces cardiac triglyceride content, to consequently improve cardiac function.⁶⁶ The complementary experiment, that is, a deletion of leptin receptors specifically in cardiomyocytes, leads to cardiovascular issues, which include impaired cardiac energy production⁶⁷ with an exacerbation in ischemic heart failure.⁶⁸ Leptin-activated STAT3 (signal transducer and activator of transcription 3) mediates the majority of leptin's physiological downstream signaling.⁶⁹ Mice harboring a deletion of STAT3 specifically in cardiac tissue are significantly more susceptible to cardiac injury following doxorubicin treatment; as a result of enhanced inflammation and cardiac fibrosis.⁷⁰ In addition, leptin conveys robust antiapoptotic effects in cardiomyocytes. In vitro, leptin treatment protects cardiomyocytes from apoptosis; potentially through its actions on improving mitochondrial function and reducing oxidative stress.^{71,72} Of note, local overexpression of leptin (by using a recombinant adenovirus expressing the leptin cDNA), prevents lipotoxic cardiomyopathy in acyl-CoA synthase transgenic mice,⁷³ potentially highlighting the antilipotoxic effects of leptin. Finally, leptin administration during reperfusion post ischemia significantly reduces infarct size.⁷⁴ Taken together, all these observations point at a substantial cardioprotective role for leptin. In addition, leptin plays an important role in regulating basal cardiac contractile function, as leptin-deficient *ob/ob* mice display impaired cardiac contractile function in ventricular myocytes.⁷⁵ Moreover, in an ex vivo system with cultured ventricular myocytes, leptin suppresses cardiac contractile function through the endothelin-1 receptor and reduced nicotinamide adenine dinucleotide phosphate oxidase-mediated pathway.^{76,77}

In contrast to the beneficial effects, in the majority of cases, leptin, particularly in the context of obesity-associated hyperleptinemia, exerts detrimental effects in cardiovascular function and promotes adverse outcomes in

cardiovascular disorders (Figure 2). In a large-scale epidemiological study, clinical observations revealed a positive correlation between hyperleptinemia and adverse cardiovascular outcomes.^{78,79} High plasma levels of leptin were shown to predict short-term occurrence of cardiac death and stroke in patients with coronary artery disease; independent of established risk factors.⁸⁰ Moreover, increasing the serum concentrations of leptin positively correlates with the total number of stenotic coronary arteries; with serum leptin levels predicting the development of arterial stiffness in patients with coronary artery disease.⁸¹ Independent of traditional risk factors (such as fasting insulin and CRP [C-reactive protein]) and metabolic abnormalities, hyperleptinemia is considered a better predictor of vascular compliance in adolescents.⁸² Furthermore, leptin is an important cardiovascular disorder marker in the obese population; this can contribute to the evaluation of metabolic risk in these individuals.⁸³ Beyond these clinical observations, rodent models have offered great insight and improved understanding toward the underlying mechanisms related to the action of leptin. Raising plasma leptin levels, by either administration of exogenous leptin or ectopic overexpression of leptin, increases arterial pressure and heart rate; this eventually leads to hypertension.⁸⁴ Cardiac leptin overexpression, in the context of acute myocardial infarction and reperfusion, potentiates myocardial remodeling and left ventricular dysfunction.⁸⁵ In contrast, the local administration of a leptin antagonist attenuates angiotensin II-induced ascending aortic aneurysms and cardiac remodeling.⁸⁶ Consistent with these observations, leptin receptor-neutralizing antibodies improve cardiac function; this offers strong evidence that endogenous leptin is a driver for cardiac hypertrophy.⁸⁷ Several mechanisms may underlie the causative effects of leptin-induced cardiovascular disorders.⁸⁸ These include induction of the the mammalian target

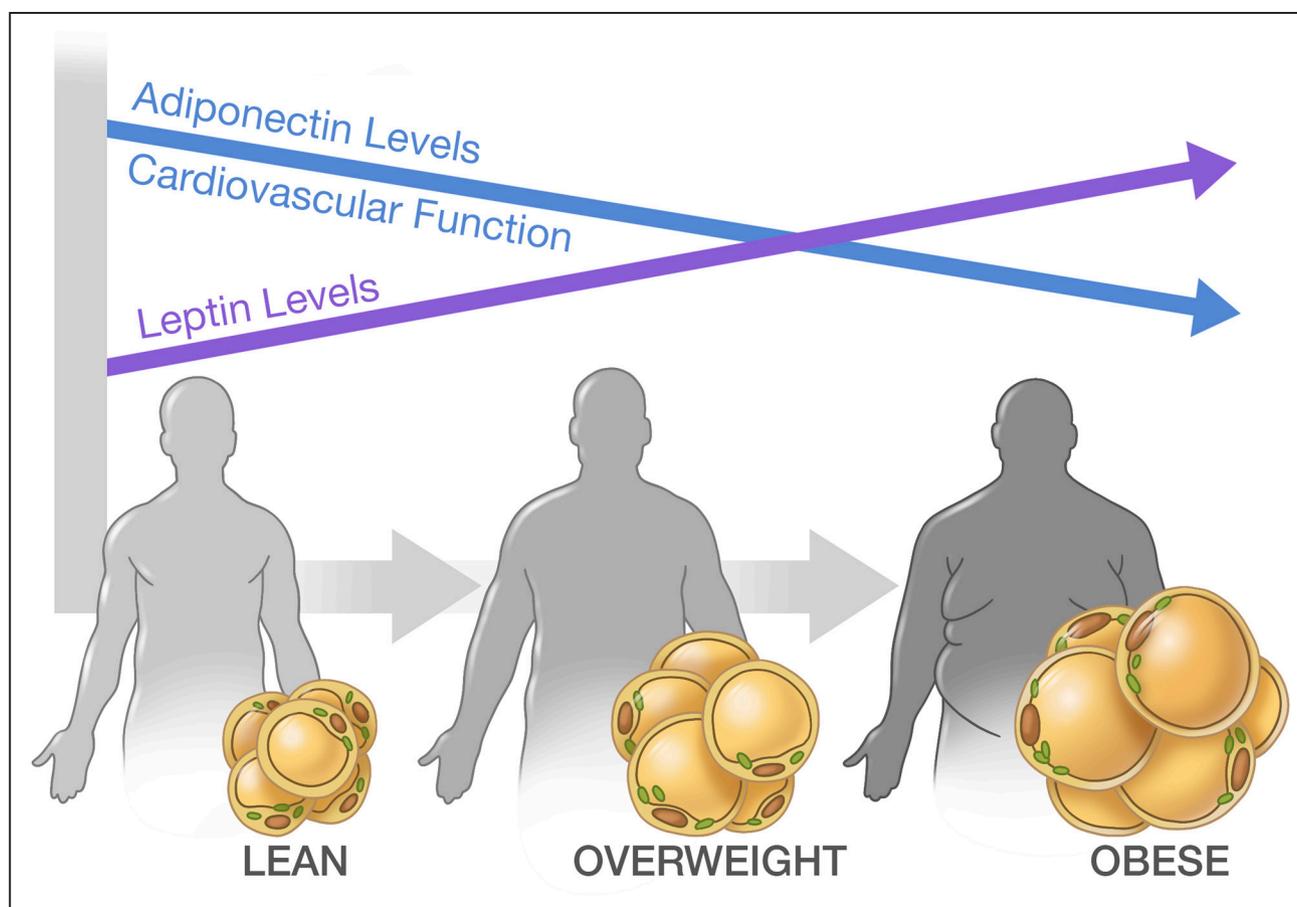


Figure 2. Relationship between body weight, fat mass, circulating leptin, and adiponectin levels with cardiovascular function.

The gradual transition from lean to overweight to obesity is associated with dramatic adipose tissue expansion. During this process, circulating leptin and adiponectin levels are altered accordingly. Low adiponectin and high leptin levels eventually negatively affect cardiovascular function.

of rapamycin pathway, activation of PPAR (peroxisome proliferator-activated receptor) α signaling, increased production of reactive oxygen species, and the activation of p38 mitogen-activated protein kinase.⁸⁸

A UNIFYING MODEL TO EXPLAIN THE PARADOXICAL EFFECTS OF LEPTIN ON CARDIOVASCULAR DYSFUNCTION

To date, there are no prevailing models to explain the seemingly paradoxical effects of leptin on cardiovascular function. Our recent observations on the effects that leptin has on body weight regulation offer a novel perspective to rationalize these paradoxical effects on the heart and vasculature. In the context of leptin sensitivity, primarily evident in young and lean mice, reducing circulating levels of leptin paradoxically results in a significant increase in food intake and body weight gain. This is, in fact, consistent with the existing models detailing the response to lower leptin concentrations. However, a different response is observed under conditions of leptin resistance. Here, a partial leptin reduction triggers a higher degree of leptin sensitivity, enhanced insulin

sensitivity, with a reduction in body weight; an unexpected and surprising response to reduced leptin levels.^{89–91} This seemingly paradoxical response to leptin reduction, in the general area of weight maintenance and energy expenditure, could also be critical for a better understanding of the paradoxical effects of leptin on cardiovascular function. Moreover, under obesogenic conditions, hyperleptinemia per se is sufficient to promote leptin resistance^{91,92}; resulting in all the other metabolic disorders frequently associated with weight gain. Thus, circulating leptin levels, in a first approximation, reflect the state of an individual's leptin sensitivity: that is, higher circulating levels of leptin equate to lower leptin sensitivity. Based on these observations, here, we put forth a new model, in which properly sustained leptin signaling, within a narrow range, is essential for normal cardiac function (Figure 3). Deficiencies in the cardiac leptin signaling pathway, as observed in *ob/ob* mice and *db/db* mice, consequently result in cardiovascular dysfunction. In contrast, chronic overactivation of the leptin signaling pathway, as observed in the diet-induced obese mice, leads to obesity-associated cardiovascular disorders. In the latter model, the beneficial impact that leptin exerts on cardiac function also follows the general rule of leptin's involvement in metabolism, that is, "less

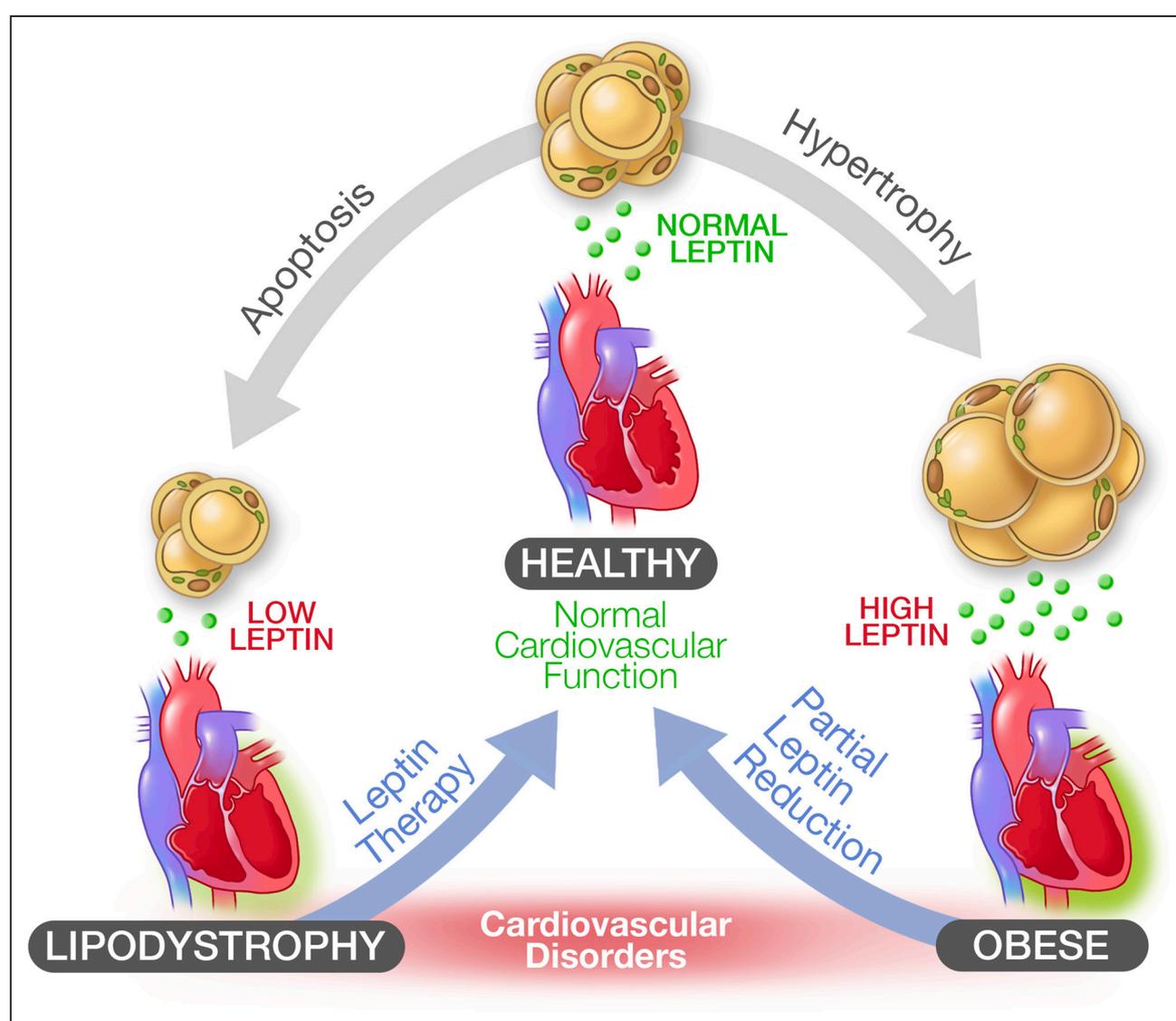


Figure 3. The relationship between the circulating levels of leptin and cardiovascular dysfunction.

For proper cardiovascular function, circulating leptin levels must be maintained within a narrow physiological range. Under conditions of lipodystrophy, caused by widespread adipose tissue apoptosis or an inability to properly develop adipose tissue, extremely low levels of circulating leptin promote cardiovascular disorders, which can be reversed by leptin therapy. Under conditions of diet-induced obesity, hyperleptinemia is a driving force for cardiovascular dysfunction due to leptin resistance, which can be restored by reducing circulating leptin levels (partial leptin reduction).

is more.⁹⁰ This provides an alternative strategy to treat obesity-associated cardiovascular disorders; essentially by lowering the circulating levels of leptin.

ADIPONECTIN

Adiponectin was first described in 1995,⁹⁵ around the same time as the initial description of leptin. Adiponectin is produced and predominately secreted by adipose tissue.⁹⁵ The adipokine exerts its beneficial effects on multiple tissues, including the heart, liver,⁹³ pancreatic β -cells,⁹⁴ the brain,⁹⁵ bone,⁹⁶ kidneys,⁹⁷ blood vessels,^{98,99} and immune cells.⁹⁹ Adiponectin in circulation exists in 3 major oligomeric multimers: a low-molecular weight trimer, a medium molecular weight hexamer, and a high

molecular weight (HMW) multimer.¹⁰⁰ HMW adiponectin represents the biologically most active form of adiponectin. In contrast to other adipokines, the circulating levels of adiponectin are inversely proportional to total fat mass.¹⁰¹ This is particularly striking for the relationship between adiponectin and leptin. Under almost all physiological conditions, these 2 adipokines are regulated in an opposite manner. High leptin generally reflects low adiponectin, and vice versa, low leptin reflects high adiponectin. For instance, obese individuals with high circulating levels of leptin, display lower levels of adiponectin. Furthermore, adiponectin secretion is predominantly determined by the quality of adipose tissue, rather than the amount of adipose tissue. Despite comparable levels of adipose tissue, metabolically healthy obesity (healthy) individuals display

higher levels of circulating adiponectin, when compared with metabolically abnormal obesity (unhealthy) individuals.¹⁰² Whether high adiponectin levels can account for all of the beneficial metabolic characteristics of metabolically healthy obesity individuals is currently not clear. However, murine models that overexpress adiponectin, do indeed exhibit a massive metabolically healthy obesity phenotype.³¹

As a pleiotropic hormone, adiponectin very robustly enhances insulin sensitivity, in addition to promoting anti-inflammatory and antifibrotic activity.^{103,104} Congenital deletion of adiponectin impairs glucose tolerance and reduces insulin sensitivity.^{105,106} This phenotype has been fully confirmed and expanded through the use of a doxycycline inducible deletion of adiponectin exclusively in mature adipose tissue.¹⁰⁷ Conversely, overexpression of adiponectin in a transgenic setting, greatly enhances insulin sensitivity, despite massive obesity.³¹ Furthermore, increasing the levels of adiponectin through exogenous administration also effectively enhances insulin sensitivity.¹⁰⁸ The beneficial effects of adiponectin are dependent on the protein binding to its receptors, AdipoR1 and AdipoR2. Deletion of AdipoR1 or AdipoR2 abolishes the beneficial effects of adiponectin.¹⁰⁹ Consistent with this observation, overexpression of AdipoR1 and AdipoR2 restores the beneficial effects of adiponectin in several tissues.¹¹⁰ Of note, an AdipoR1 and AdipoR2 agonist (AdipoRon), ameliorates diabetes in a genetically obese rodent model (*db/db* mice) and prolongs the life-span of *db/db* mice during high-fat diet feeding.¹¹¹ Our own studies established that the potent ceramide-reducing effect of adiponectin relies on the ceramidase domain contained within the AdipoR1 and AdipoR2 receptors.¹¹⁰

THE PARADOXICAL EFFECT OF ADIPONECTIN ON CARDIOVASCULAR DYSFUNCTION

Given its robust effects on inflammation and fibrosis, we would predict that adiponectin has a protective role against cardiovascular disorders. There is no doubt that low levels of adiponectin are tightly associated with the increased prevalence of obesity-linked cardiovascular disorders, including ischemic heart disease and peripheral artery disease (Figure 2). However, the situation is more complex in the setting of higher circulating adiponectin levels. In some cases, higher circulating levels of adiponectin are associated with a better outcome for cardiovascular events. Conversely in other cases, higher levels of adiponectin are associated with no beneficial effects, or even detrimental effects, such as increasing mortality rate. At times, this is referred to as the adiponectin paradox (Figure 1). This phenomenon was first observed in the context of cardiovascular and kidney disease. It has also been evident in a subset of elderly patients with type 2 diabetes.^{112,113}

Given that adiponectin levels are inversely correlated with fat mass, obesity-associated hypoadiponectinemia may serve as a bridge between obesity and cardiovascular disorders. Numerous human studies have provided firm evidence that hypoadiponectinemia is associated with adverse cardiovascular events. For instance, in patients with coronary artery disease, the ratio of HMW adiponectin per total adiponectin is significantly lower, while the trimeric form is significantly higher. Consistent with these observations, weight loss in obese individuals increases the HMW form of adiponectin, while reducing the hexameric and trimeric forms.¹¹⁴ In agreement with clinical data, cell-culture based studies and murine model systems further provide a clear picture that adiponectin, particularly the HMW form, is beneficial for cardiovascular function.

DISSECTING THE PARADOXICAL EFFECT OF ADIPONECTIN

To delineate the mechanisms that underlie the paradoxical effects of adiponectin in cardiovascular mortality, we turn our attention to the source of circulating adiponectin, in the context of cardiovascular dysfunction. In theory, the circulating levels of adiponectin are determined by the intricate balance between production and clearance. Adipose tissue is the predominant source for circulating adiponectin. Cardiomyocytes can also produce small amounts of adiponectin, which exert local autocrine or paracrine effects, however, do not significantly contribute to circulating levels.¹¹⁵ As such, adipose tissue is the primary source of adiponectin released into circulation. Beyond its origin, the quality, not quantity, of adipose tissue determines the rate of adiponectin release. In patients with severe cardiovascular disorders, the quality of adipose tissue is largely compromised, as reflected by increased adipose tissue inflammation.¹¹⁶ Therefore, increased levels of adiponectin may be attributed to delayed clearance, rather than production. Adiponectin is rapidly cleared with a half-life of ≈ 75 minutes under normal physiological conditions, primarily by the liver¹¹⁷ and to a much lower extent, the kidney. High adiponectin levels are detected during chronic liver disease, and further, correlate with inflammation and liver damage; reflecting a delayed clearance rate.¹¹⁸ Additionally, diet-induced obese mice, or *db/db* mice, exhibit a much slower rate of adiponectin clearance. This points to a liver-mediated delay in adiponectin clearance as the primary cause for the higher levels of adiponectin evident in patients with cardiovascular disorders. Consistent with this observation, the adiponectin paradox frequently occurs in patients with both cardiovascular disorders and other metabolic disorders, such as liver or kidney disease. Beyond synthesis and clearance, other variables, such as race, sex, age, smoking, body mass index (BMI), and drug regimen history, can determine the levels of adiponectin in circulation. The circulating levels of adiponectin are increased with age; a phenomenon that is more

pronounced in men than in women.¹¹⁹ Moreover, BMI is a strong correlate that determines adiponectin levels, that is, subjects with a lower BMI tend to exhibit higher circulating levels of adiponectin. Furthermore, certain pharmacological interventions that directly target adipose tissue, have been shown to regulate adiponectin secretion. For instance, the PPAR γ agonist rosiglitazone, a classical drug used to treat type 2 diabetic patients, increases adiponectin secretion.¹²⁰ Furthermore, cardiomyocytes may also contribute to the increase in the rate of adiponectin production. However, the physiological relevance of the latter remains to be evaluated, through use of a murine model of cardiomyocyte-specific elimination of adiponectin. Taken together, these confounding factors all contribute to the increased levels of circulating adiponectin evident in patients with cardiovascular dysfunction. Based on these observations, the elevated levels of adiponectin may be a secondary consequence, rather than a primary driver of cardiovascular dysfunction.

Another unanswered question is whether the high levels of adiponectin, in the context of the adiponectin paradox, reflect fully functional material? In some population-based studies, that is, in elderly individuals with a cardiovascular disorder, there is a significant correlation between the total levels of adiponectin with a higher mortality rate. The correlation between HMW adiponectin and increased mortality is, however, less straightforward.^{121,122} Given HMW adiponectin accounts for much of the protective effects in the heart, this may raise the issue as to whether adiponectin is in a functional configuration in these settings. So far, no attempts have been made to directly verify this. Utilizing more functional readouts to confirm the role of HMW adiponectin in the heart, in the context of the adiponectin paradox, are required to fully understand the reasons for the disproportionately high levels of the protein in circulation.

As the physiological role of adiponectin is well preserved from mouse to human, observations made in the context of mouse models generally translate to significant implications for clinical studies. The adiponectin paradox is primarily observed in human prospective studies. Surprisingly, we have not observed this paradox in any rodent models. The presence of high levels of adiponectin in individuals with a severe cardiovascular disorder may be merely a compensatory response in attempt to restore heart function; rather than reflecting a detrimental role to accelerate the disease progression. Currently, no attempt has been made to reduce adiponectin levels in patients with cardiovascular disorders, to support the idea that adiponectin directly contributes to disease progression on the basis of its upregulation. Rodent data strongly argues for the beneficial aspects of adiponectin. The inducible deletion of adiponectin in adult mice produces a strong phenotype; this includes profound insulin resistance and inflammation, severely impaired glucose tolerance and delayed lipid clearance.¹⁰⁷ While high adiponectin levels, on

the contrary, as achieved by transgenic overexpression or exogenous administration, produces substantial improvement in cardiovascular function.¹²³ Furthermore, activation of the adiponectin receptors (AdipoR1 and AdipoR2), by using AdipoRon, alleviates the diabetic phenotype in genetically obese rodent models, that is, in the *db/db* mouse; a mouse model that exhibits severe cardiovascular dysfunction. At the cellular level, in vitro studies using cells or primary cardiomyocytes isolated from heart tissues of fetal or adult rodents, have been used to show a protective effect of adiponectin on cardiomyocyte function.

USING LEPTIN AND ADIPONECTIN AS A STRATEGY TO PREVENT OR TREAT CARDIOVASCULAR DYSFUNCTION

Proper leptin signaling is necessary and indispensable to maintain optimal performance of the heart (Figure 2). Leptin insufficiency, primarily observed in lipodystrophy and leptin-deficient mouse models, is associated with cardiovascular disorders.^{75,124–126} In contrast, hyperleptinemia, frequently observed in diet-induced obesity, also results in cardiovascular disorders and increased mortality rates. As such, targeting a leptin-based therapy to treat cardiovascular disorders, should focus on the circulating levels of leptin. In the setting of complete leptin insufficiency, leptin therapy (by elevating leptin concentrations to physiological levels) is adequate to prevent or reverse cardiovascular disorders (Figure 3). Successful application of metreleptin (a recombinant form of leptin) to treat patients with lipodystrophy has been well established. More specifically, leptin therapy in these lipodystrophic individuals very effectively normalizes glucose tolerance and insulin sensitivity, to improve liver and heart function.¹²⁷ However, in the context of hyperleptinemia, leptin therapy is largely ineffective. Rather, a partial leptin reduction strategy, by reducing the circulating levels of leptin to achieve a normal systemic range, shows great promise in treating obesity and its associated cardiovascular disorders (Figure 3). Of note in this context, this approach does not require a sophisticated titration to normalize leptin levels. Based on our experience in the area of metabolism, we identified that there is a wide therapeutic range available for leptin neutralization therapy. Any significant reductions in leptin trigger beneficial effects, that is, leptin levels can be very significantly lowered, but the benefits of this reduction are still preserved.

PARTIAL LEPTIN REDUCTION: A NOVEL STRATEGY FOR THE TREATMENT OF HYPERLEPTINEMIA-ASSOCIATED CARDIOVASCULAR DYSFUNCTION

Recently, we demonstrated that hyperleptinemia per se is sufficient to promote leptin resistance. High leptin

levels serve as a driving force for obesity and its associated metabolic disorders. This offered the possibility that reducing leptin levels in circulation lends itself as an effective strategy to treat obesity and its metabolic consequences.⁹⁰ We have already achieved this, essentially by using genetic mouse models, as well as through leptin neutralizing antibodies. Partial leptin reduction restores the physiological role of leptin in reducing food intake and enhancing energy expenditure, which leads to significant weight loss and antidiabetic effects. Based on these findings, we propose that partial leptin reduction will also alleviate the pathogenesis of cardiovascular dysfunction, to effectively improve the outcome of a cardiovascular event.

The effectiveness of partial leptin reduction, in the context of hyperleptinemia, has been indirectly supported by many observations. Lifestyle alterations and pharmacological interventions that demonstrate cardiovascular improvement are associated with a partial reduction in the circulating levels of leptin. For instance, long-term caloric restriction preserves cardiac contractile function, to improve cardiomyocyte function and reduce cardiac remodeling.^{128,129} Caloric restriction also effectively lowers the circulating levels of leptin.^{130,131} High-intensity training has also been shown to cause partial leptin reduction, concomitant with improved cardiovascular function.¹³² From a pharmacological perspective, the glucagon-like peptide-1 analog, liraglutide, greatly reduces cardiovascular events and mortality rate; this is associated with reduced levels of leptin.¹³³ Moreover, inhibitors targeting the SGLT-2 (sodium-glucose-linked transporter-2), have emerged as one of the most powerful classes of cardiovascular drugs in recent years.¹³⁴ However, little is known about the underlying mechanisms that drive their cardiovascular benefits. Reduced levels of leptin, in the context of SGLT-2 inhibition, could be one of the highly likely mechanisms underlying this phenomenon.¹³⁵ Hyperleptinemia results in sodium retention and plasma volume expansion; this can activate cardiac and renal inflammation and fibrosis. Furthermore, leptin-mediated neuro-hormonal activation appears to increase the expression of SGLT-2 in the renal tubule.¹³⁶ Other potent cardiovascular interventions, such as the angiotensin-converting enzyme inhibitor perindopril,¹³⁷ or metformin, or statins, have direct effects on adipocytes, specifically on white adipose tissue, to decrease leptin expression.^{138,139} Finally, CB1 (cannabinoid receptor 1) antagonists have also been used as an additional strategy to lower the circulating levels of leptin.^{140,141} Collectively, these known effects of leptin correlate well with cardiovascular benefits, suggesting the possibility to directly target leptin reduction for the treatment of cardiovascular disorders. We therefore propose that reducing the circulating levels of leptin, in the context of hyperleptinemia, can directly lead to cardiovascular improvement. Leptin neutralizing

antibodies that effectively lower the circulating levels of leptin,⁹¹ display great promise in inducing weight loss, in addition to exerting antifibrotic and insulin sensitizing effects. To date, however, the effects of such antibody-based approaches on the cardiovascular system are still awaiting further investigation.

ADIPONECTIN THERAPY IN THE PREVENTION OF CARDIOVASCULAR DYSFUNCTION

In individuals that harbor lower circulating levels of adiponectin, identifying means to elevate adiponectin levels to a physiological range, still holds great promise for the treatment of cardiovascular dysfunction. A subset of approved pharmacological interventions that display beneficial effects on cardiovascular disorders, are also associated with increased circulating levels of adiponectin. For example, the use of PPAR γ agonists results in a robust increase in adiponectin levels.¹⁴² As an alternative means, rather than increasing the circulating levels of adiponectin, enhancing adiponectin signaling could also serve as an additional strategy to treat cardiovascular disorders. The identification of a small molecule agonist of the adiponectin receptors AdipoR1 and AdipoR2 (referred to as AdipoRon), has generated much interest in the identification of additional ligands targeted towards improvements in cardiovascular disorder. More specifically, several rodent models that enhance adiponectin receptor signaling through AdipoRon, have provided proof-of-principle that this approach may constitute an effective promising therapy to treat cardiovascular dysfunction.¹⁴³ As an additional binding partner, T-cadherin (that also binds to adiponectin) has also garnered attention in the treatment of cardiovascular disorder,¹⁴⁴ and as such, some aspects of the cardioprotective effects that adiponectin elicits may be mediated through this additional receptor.¹⁴⁵ In light of this, the adiponectin/T-cadherin complex has been shown to provide cardiovascular protection by enhancing exosomal production and release; secreting cell-toxic products from specific cell types, that is, cells within the vasculature. In this respect, future studies in the area of T-cadherin signaling as a therapeutic intervention to treat cardiovascular disorders should prove illuminating.

INCREASING THE ADIPONECTIN/LEPTIN RATIO: AN EMERGING STRATEGY TO TREAT CARDIOVASCULAR DYSFUNCTION

Instead of individually targeting adiponectin and leptin, interventions that directly act on both axis separately, to increase the adiponectin to leptin ratio have recently garnered attention.¹⁴⁶⁻¹⁴⁹ In chow-fed mice, the average adiponectin levels are approximately 15 to 20 $\mu\text{g/mL}$,

while the circulating leptin levels are approximately 5 to 10 ng/mL; as such, the calculated ratio is between 1 and 4;⁹¹ a ratio positively associated with metabolic health and reduced cardiovascular disorder. However, in diet-induced obese mice, this ratio is greatly reduced, as a result of unhealthy adipose tissue expansion, leading to dysfunctional adipose tissue and cardiovascular disorders, which are characterized by increased systemic inflammation and tissue fibrosis.^{20,21,150} In diet-induced obese mice, adiponectin levels drop to 10–15 µg/mL, and leptin levels are greatly increased to 50–150 ng/mL, and thus the calculated ratio is much lower than one.⁹¹ In clinical settings, the ratio of adiponectin to leptin is a more predictive and reliable biomarker for several metabolic disorders, such as insulin resistance, type 2 diabetes, hypertension, and cardiovascular disorders.¹⁵¹ Therefore, in obese patients with cardiovascular dysfunction, an ideal treatment would be to combine the beneficial effects of both adiponectin therapy and partial leptin reduction. Weight loss is associated with a significantly elevated ratio, as progressive weight loss elevates adiponectin levels and reduces leptin levels.¹⁵² Thus, pharmacological interventions and bariatric surgery that induce substantial weight loss, could serve as a strategy to elevate the ratio.^{153–156} However, it is still unclear whether the cardiovascular beneficial effects of weight loss are directly derived from an elevated ratio of adiponectin to leptin. Further experiments are definitely warranted to confirm this causing effect.

Independent of significant weight loss, simpler treatments that aim to simultaneously elevate adiponectin and reduce leptin levels are still under development. The recently developed leptin neutralizing antibody, together with PPAR γ agonists that lead to long-lasting adiponectin increases, may be an excellent combination therapy to achieve an elevated ratio. The efficacy and efficiency of this combination therapy in reversing cardiovascular disorders is yet to be determined. Based on the positive outcomes of monotherapy of leptin neutralization, a much better outcome in treating obesity-associated cardiovascular disorders could be expected. In addition, the existence of multiple mouse models, including doxycycline inducible leptin transgenic mice, adiponectin overexpression mice, inducible leptin and adiponectin KO (knockout) mice with tissue specific Cre expression,¹⁵⁷ will allow us to better examine the cause and effect relationship of the ratio of adiponectin to leptin in cardiovascular disorders.

CONCLUSIONS AND PERSPECTIVE

The pathogenesis of cardiovascular dysfunction is highly complex. Two of the most widely studied adipokines, adiponectin and leptin, are important players in determining cardiovascular disorder progression and outcome. Essentially, both adipokines are required for

proper cardiovascular function. Impaired leptin or adiponectin signaling, due to lipodystrophy or genetic mutations, results in an adverse outcome of cardiovascular dysfunction. On the contrary, an oversupply of leptin or adiponectin in circulation, can directly exert a negative cardiovascular impact; whereas the paradoxical increase of adiponectin in this context, is likely a reflection of a compensatory response. A combined approach aimed at restoring normal physiological levels of both adipokines is highly likely to elicit a positive cardiovascular outcome. Here, we propose that an increased ratio of adiponectin to leptin can emerge as a highly promising and aspirational therapeutic goal.

ARTICLE INFORMATION

Affiliations

Touchstone Diabetes Center, Department of Internal Medicine (S.Z., C.M.K., P.E.S.) and Department of Cell Biology (P.E.S.), The University of Texas Southwestern Medical Center, Dallas.

Sources of Funding

The authors are supported by US National Institutes of Health (NIH) grants R01-DK55758, RC2-DK118620, P01-DK088761, R01-DK099110, and P01-AG051459 (P.E. Scherer); S. Zhao is supported by a Post-Doctoral Fellowship from Fondation de Recherche Santé Québec (FRQS). C.M. Kusminski is supported by SRA201808-0004 from Amgen and SRA201909-0007 from Eli Lilly.

Acknowledgments

We would like to thank Richard Howdy from Visually Medical for help with the graphics.

Disclosures

None.

REFERENCES

1. Jaacks LM, Vandevijvere S, Pan A, McGowan CJ, Wallace C, Imamura F, Mozaffarian D, Swinburn B, Ezzati M. The obesity transition: stages of the global epidemic. *Lancet Diabetes Endocrinol*. 2019;7:231–240. doi: 10.1016/S2213-8587(19)30026-9
2. Kusminski CM, Bickel PE, Scherer PE. Targeting adipose tissue in the treatment of obesity-associated diabetes. *Nat Rev Drug Discov*. 2016;15:639–660. doi: 10.1038/nrd.2016.75
3. Mc Namara K, Alzubaidi H, Jackson JK. Cardiovascular disease as a leading cause of death: how are pharmacists getting involved? *Integr Pharm Res Pract*. 2019;8:1–11. doi: 10.2147/IPRPS133088
4. Lycett K, Juonala M, Magnussen CG, Norrish D, Mensah FK, Liu R, Clifford SA, Carlin JB, Olds T, Saffery R, et al. Body mass index from early to late childhood and cardiometabolic measurements at 11 to 12 years. *Pediatrics*. 2020;146:e20193666. doi: 10.1542/peds.2019-3666
5. Koliaki C, Liatis S, Kokkinos A. Obesity and cardiovascular disease: revisiting an old relationship. *Metabolism*. 2019;92:98–107. doi: 10.1016/j.metabol.2018.10.011
6. Kratz M, Baars T, Guyenet S. The relationship between high-fat dairy consumption and obesity, cardiovascular, and metabolic disease. *Eur J Nutr*. 2013;52:1–24. doi: 10.1007/s00394-012-0418-1
7. Lee MJ, Wu Y, Fried SK. Adipose tissue heterogeneity: implication of depot differences in adipose tissue for obesity complications. *Mol Aspects Med*. 2013;34:1–11. doi: 10.1016/j.mam.2012.10.001
8. Zhu Y, Gao Y, Tao C, Shao M, Zhao S, Huang W, Yao T, Johnson JA, Liu T, Cypess AM, et al. Connexin 43 mediates white adipose tissue beiging by facilitating the propagation of sympathetic neuronal signals. *Cell Metab*. 2016;24:420–433. doi: 10.1016/j.cmet.2016.08.005
9. Zhang Z, Shao M, Hepler C, Zi Z, Zhao S, An YA, Zhu Y, Ghaheri AL, Wang MY, Li N, et al. Dermal adipose tissue has high plasticity and undergoes reversible dedifferentiation in mice. *J Clin Invest*. 2019;129:5327–5342. doi: 10.1172/JCI130239

10. Zhang F, Hao G, Shao M, Nham K, An Y, Wang Q, Zhu Y, Kusminski CM, Hassan G, Gupta RK, et al. An adipose tissue atlas: an image-guided identification of human-like BAT and beige depots in rodents. *Cell Metab*. 2018;27:252.e3–262.e3. doi: 10.1016/j.cmet.2017.12.004
11. Lee JJ, Pedley A, Hoffmann U, Massaro JM, O'Donnell CJ, Benjamin EJ, Long MT. Longitudinal associations of pericardial and intrathoracic fat with progression of coronary artery calcium (from the Framingham Heart Study). *Am J Cardiol*. 2018;121:162–167. doi: 10.1016/j.amjcard.2017.10.006
12. Hall JE, do Carmo JM, da Silva AA, Wang Z, Hall ME. Obesity-induced hypertension: interaction of neurohumoral and renal mechanisms. *Circ Res*. 2015;116:991–1006. doi: 10.1161/CIRCRESAHA.116.305697
13. Hepler C, Shan B, Zhang Q, Henry GH, Shao M, Vishvanath L, Ghaben AL, Mobley AB, Strand D, Hon GC, et al. Identification of functionally distinct fibro-inflammatory and adipogenic stromal subpopulations in visceral adipose tissue of adult mice. *Elife*. 2018;7:e39636.
14. Schwalie PC, Dong H, Zachara M, Russeil J, Alpern D, Akchiche N, Caprara C, Sun W, Schlaudraff KU, Soldati G, et al. A stromal cell population that inhibits adipogenesis in mammalian fat depots. *Nature*. 2018;559:103–108. doi: 10.1038/s41586-018-0226-8
15. Zhao S, Mugabo Y, Ballentine G, Attane C, Iglesias J, Poursharifi P, Zhang D, Nguyen TA, Erb H, Prentki R, et al. α/β -hydrolase domain 6 deletion induces adipose browning and prevents obesity and type 2 diabetes. *Cell Rep*. 2016;14:2872–2888. doi: 10.1016/j.celrep.2016.02.076
16. Planat-Benard V, Silvestre JS, Cousin B, André M, Nibbelink M, Tamarat R, Clergue M, Manneville C, Saillan-Barreau C, Duriez M, et al. Plasticity of human adipose lineage cells toward endothelial cells: physiological and therapeutic perspectives. *Circulation*. 2004;109:656–663. doi: 10.1161/01.CIR.0000114522.38265.61
17. Sun X, Lin J, Zhang Y, Kang S, Belkin N, Wara AK, Icli B, Hamburg NM, Li D, Feinberg MW. MicroRNA-181b improves glucose homeostasis and insulin sensitivity by regulating endothelial function in white adipose tissue. *Circ Res*. 2016;118:810–821. doi: 10.1161/CIRCRESAHA.115.308166
18. Sun K, Kusminski CM, Scherer PE. Adipose tissue remodeling and obesity. *J Clin Invest*. 2011;121:2094–2101. doi: 10.1172/JCI45887
19. Crewe C, An YA, Scherer PE. The ominous triad of adipose tissue dysfunction: inflammation, fibrosis, and impaired angiogenesis. *J Clin Invest*. 2017;127:74–82. doi: 10.1172/JCI88883
20. Reilly SM, Saltiel AR. Adapting to obesity with adipose tissue inflammation. *Nat Rev Endocrinol*. 2017;13:633–643. doi: 10.1038/nrendo.2017.90
21. Zhu Q, An YA, Kim M, Zhang Z, Zhao S, Zhu Y, Asterholm IW, Kusminski CM, Scherer PE. Suppressing adipocyte inflammation promotes insulin resistance in mice. *Mol Metab*. 2020;39:101010. doi: 10.1016/j.molmet.2020.101010
22. Ghaben AL, Scherer PE. Adipogenesis and metabolic health. *Nat Rev Mol Cell Biol*. 2019;20:242–258. doi: 10.1038/s41580-018-0093-z
23. Bays HE, González-Campoy JM, Bray GA, Kitabchi AE, Bergman DA, Schorr AB, Rodbard HW, Henry RR. Pathogenic potential of adipose tissue and metabolic consequences of adipocyte hypertrophy and increased visceral adiposity. *Expert Rev Cardiovasc Ther*. 2008;6:343–368. doi: 10.1586/14779072.6.3.343
24. Shao M, Vishvanath L, Busbuso NC, Hepler C, Shan B, Sharma AX, Chen S, Yu X, An YA, Zhu Y, et al. De novo adipocyte differentiation from Pdgfr β -preadipocytes protects against pathologic visceral adipose expansion in obesity. *Nat Commun*. 2018;9:890. doi: 10.1038/s41467-018-03196-x
25. Wang QA, Tao C, Gupta RK, Scherer PE. Tracking adipogenesis during white adipose tissue development, expansion and regeneration. *Nat Med*. 2013;19:1338–1344. doi: 10.1038/nm.3324
26. Kopelman PG. Obesity as a medical problem. *Nature*. 2000;404:635–643. doi: 10.1038/35007508
27. Seip M, Trygstad O. Generalized lipodystrophy, congenital and acquired (lipoatrophy). *Acta Paediatr Suppl*. 1996;413:2–28. doi: 10.1111/j.1651-2227.1996.tb14262.x
28. Karelis AD. Metabolically healthy but obese individuals. *Lancet*. 2008;372:1281–1283. doi: 10.1016/S0140-6736(08)61531-7
29. Smith GI, Mittendorfer B, Klein S. Metabolically healthy obesity: facts and fantasies. *J Clin Invest*. 2019;129:3978–3989. doi: 10.1172/JCI129186
30. Morley TS, Xia JY, Scherer PE. Selective enhancement of insulin sensitivity in the mature adipocyte is sufficient for systemic metabolic improvements. *Nat Commun*. 2015;6:7906. doi: 10.1038/ncomms8906
31. Kim JY, van de Wall E, Laplante M, Azzara A, Trujillo ME, Hofmann SM, Schraw T, Durand JL, Li H, Li G, et al. Obesity-associated improvements in metabolic profile through expansion of adipose tissue. *J Clin Invest*. 2007;117:2621–2637. doi: 10.1172/JCI31021
32. Kusminski CM, Holland WL, Sun K, Park J, Spurgin SB, Lin Y, Askew GR, Simcox JA, McClain DA, Li C, et al. MitoNEET-driven alterations in adipocyte mitochondrial activity reveal a crucial adaptive process that preserves insulin sensitivity in obesity. *Nat Med*. 2012;18:1539–1549. doi: 10.1038/nm.2899
33. Fabbrini E, Yoshino J, Yoshino M, Magkos F, Tiemann Luecking C, Samovski D, Fraterrigo G, Okunade AL, Patterson BW, Klein S. Metabolically normal obese people are protected from adverse effects following weight gain. *J Clin Invest*. 2015;125:787–795. doi: 10.1172/JCI78425
34. Stefan N, Häring HU, Hu FB, Schulze MB. Metabolically healthy obesity: epidemiology, mechanisms, and clinical implications. *Lancet Diabetes Endocrinol*. 2013;1:152–162. doi: 10.1016/S2213-8587(13)70062-7
35. Scherer PE, Williams S, Fogliano M, Baldini G, Lodish HF. A novel serum protein similar to C1q, produced exclusively in adipocytes. *J Biol Chem*. 1995;270:26746–26749. doi: 10.1074/jbc.270.45.26746
36. Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature*. 1994;372:425–432. doi: 10.1038/372425a0
37. Funcke JB, Scherer PE. Beyond adiponectin and leptin: adipose tissue-derived mediators of inter-organ communication. *J Lipid Res*. 2019;60:1648–1684. doi: 10.1194/jlr.R094060
38. Caron A, Lee S, Elmquist JK, Gautron L. Leptin and brain-adipose cross-talks. *Nat Rev Neurosci*. 2018;19:153–165. doi: 10.1038/nrn.2018.7
39. Wang J, Liu R, Hawkins M, Barzilai N, Rossetti L. A nutrient-sensing pathway regulates leptin gene expression in muscle and fat. *Nature*. 1998;393:684–688. doi: 10.1038/31474
40. Cammisotto PG, Renaud C, Gingras D, Delvin E, Levy E, Bédard M. Endocrine and exocrine secretion of leptin by the gastric mucosa. *J Histochem Cytochem*. 2005;53:851–860. doi: 10.1369/jhc.5A6620.2005
41. Ashworth CJ, Hoggard N, Thomas L, Mercer JG, Wallace JM, Lea RG. Placental leptin. *Rev Reprod*. 2000;5:18–24. doi: 10.1530/ror.00050018
42. Jayachandran T, Srinivasan B, Padmanabhan S. Salivary leptin levels in normal weight and overweight individuals and their correlation with orthodontic tooth movement. *Angle Orthod*. 2017;87:739–744. doi: 10.2319/120216-869.1
43. Odle AK, Haney A, Allensworth-James M, Akhter N, Childs GV. Adipocyte versus pituitary leptin in the regulation of pituitary hormones: somatotropes develop normally in the absence of circulating leptin. *Endocrinology*. 2014;155:4316–4328. doi: 10.1210/en.2014-1172
44. Blum WF, Englaro P, Hanitsch S, Juul A, Hertel NT, Müller J, Skakkebaek NE, Heiman ML, Birkett M, Attanasio AM, et al. Plasma leptin levels in healthy children and adolescents: dependence on body mass index, body fat mass, gender, pubertal stage, and testosterone. *J Clin Endocrinol Metab*. 1997;82:2904–2910. doi: 10.1210/jcem.82.9.4251
45. Dallongeville J, Fruchart JC, Auwerx J. Leptin, a pleiotropic hormone: physiology, pharmacology, and strategies for discovery of leptin modulators. *J Med Chem*. 1998;41:5337–5352. doi: 10.1021/jm9802867
46. Farooqi IS, O'Rahilly S. Leptin: a pivotal regulator of human energy homeostasis. *Am J Clin Nutr*. 2009;89:980S–984S. doi: 10.3945/ajcn.2008.26788C
47. Fischer AW, Cannon B, Nedergaard J. Leptin: is it thermogenic? *Endocr Rev*. 2020;41:232–260.
48. Shek EW, Brands MW, Hall JE. Chronic leptin infusion increases arterial pressure. *Hypertension*. 1998;31:409–414. doi: 10.1161/01.hyp.31.1.409
49. Francisco V, Pino J, Campos-Cabaleiro V, Ruiz-Fernández C, Mera A, Gonzalez-Gay MA, Gómez R, Gualillo O. Obesity, fat mass and immune system: role for leptin. *Front Physiol*. 2018;9:640. doi: 10.3389/fphys.2018.00640
50. Rosenbaum M, Leibel RL. 20 years of leptin: role of leptin in energy homeostasis in humans. *J Endocrinol*. 2014;223:T83–T96. doi: 10.1530/JOE-14-0358
51. Elmquist JK, Bjorbaek C, Ahima RS, Flier JS, Saper CB. Distributions of leptin receptor mRNA isoforms in the rat brain. *J Comp Neurol*. 1998;395:535–547.
52. Flier JS. Starvation in the midst of plenty: reflections on the history and biology of insulin and leptin. *Endocr Rev*. 2019;40:1–16. doi: 10.1210/er.2018-00179
53. Luheshi GN, Gardner JD, Rushforth DA, Loudon AS, Rothwell NJ. Leptin actions on food intake and body temperature are mediated by IL-1. *Proc Natl Acad Sci USA*. 1999;96:7047–7052. doi: 10.1073/pnas.96.12.7047
54. Ahima RS, Prabakaran D, Mantzoros C, Qu D, Lowell B, Maratos-Flier E, Flier JS. Role of leptin in the neuroendocrine response to fasting. *Nature*. 1996;382:250–252. doi: 10.1038/382250a0
55. Sinha MK, Sturis J, Ohannesian J, Magosin S, Stephens T, Heiman ML, Polonsky KS, Caro JF. Ultradian oscillations of leptin secretion in humans. *Biochem Biophys Res Commun*. 1996;228:733–738. doi: 10.1006/bbrc.1996.1724

56. Larsen MA, Isaksen VT, Paulssen EJ, Goll R, Florholmen JR. Postprandial leptin and adiponectin in response to sugar and fat in obese and normal weight individuals. *Endocrine*. 2019;66:517–525. doi: 10.1007/s12020-019-02102-9
57. Pan WW, Myers MG Jr. Leptin and the maintenance of elevated body weight. *Nat Rev Neurosci*. 2018;19:95–105. doi: 10.1038/nrn.2017.168
58. Myers MG, Cowley MA, Münzberg H. Mechanisms of leptin action and leptin resistance. *Annu Rev Physiol*. 2008;70:537–556. doi: 10.1146/annurev.physiol.70.113006.100707
59. Ren J. Leptin and hyperleptinemia - from friend to foe for cardiovascular function. *J Endocrinol*. 2004;181:1–10. doi: 10.1677/joe.0.1810001
60. Martínez-Martínez E, Jurado-López R, Cervantes-Escalera P, Cachofeiro V, Miana M. Leptin, a mediator of cardiac damage associated with obesity. *Horm Mol Biol Clin Invest*. 2014;18:3–14. doi: 10.1515/hmbci-2013-0060
61. Unger RH. Hyperleptinemia: protecting the heart from lipid overload. *Hypertension*. 2005;45:1031–1034. doi: 10.1161/01.HYP0000165683.09053.02
62. Purdham DM, Zou MX, Rajapurohitam V, Karmazyn M. Rat heart is a site of leptin production and action. *Am J Physiol Heart Circ Physiol*. 2004;287:H2877–H2884. doi: 10.1152/ajpheart.00499.2004
63. Ahiane BO, Smith W, Lammertyn L, Schutte AE. Leptin and the vasculature in young adults: The African-PREDICT study. *Eur J Clin Invest*. 2019;49:e13039. doi: 10.1111/eci.13039
64. Mori J, Patel VB, Abo Alrob O, Basu R, Altamimi T, Desaulniers J, Wagg CS, Kassiri Z, Lopaschuk GD, Oudit GY. Angiotensin 1-7 ameliorates diabetic cardiomyopathy and diastolic dysfunction in db/db mice by reducing lipotoxicity and inflammation. *Circ Heart Fail*. 2014;7:327–339. doi: 10.1161/CIRCHEARTFAILURE.113.000672
65. Barouch LA, Berkowitz DE, Harrison RW, O'Donnell CP, Hare JM. Disruption of leptin signaling contributes to cardiac hypertrophy independently of body weight in mice. *Circulation*. 2003;108:754–759. doi: 10.1161/01.CIR.0000083716.82622.FD
66. Hall ME, Mearns MW, Hall JE, Stec DE. Rescue of cardiac leptin receptors in db/db mice prevents myocardial triglyceride accumulation. *Am J Physiol Endocrinol Metab*. 2014;307:E316–E325. doi: 10.1152/ajpendo.00005.2014
67. Hall ME, Smith G, Hall JE, Stec DE. Cardiomyocyte-specific deletion of leptin receptors causes lethal heart failure in Cre-recombinase-mediated cardiotoxicity. *Am J Physiol Regul Integr Comp Physiol*. 2012;303:R1241–R1250. doi: 10.1152/ajpregu.00292.2012
68. McGaffin KR, Witham WG, Yester KA, Romano LC, O'Doherty RM, McTiernan CF, O'Donnell CP. Cardiac-specific leptin receptor deletion exacerbates ischaemic heart failure in mice. *Cardiovasc Res*. 2011;89:60–71. doi: 10.1093/cvr/cvq288
69. Bates SH, Stearns WH, Dundon TA, Schubert M, Tso AW, Wang Y, Banks AS, Lavery HJ, Haq AK, Maratos-Flier E, et al. STAT3 signalling is required for leptin regulation of energy balance but not reproduction. *Nature*. 2003;421:856–859. doi: 10.1038/nature01388
70. Jacoby JJ, Kalinowski A, Liu MG, Zhang SS, Gao Q, Chai GX, Ji L, Iwamoto Y, Li E, Schneider M, et al. Cardiomyocyte-restricted knockout of STAT3 results in higher sensitivity to inflammation, cardiac fibrosis, and heart failure with advanced age. *Proc Natl Acad Sci USA*. 2003;100:12929–12934. doi: 10.1073/pnas.2134694100
71. Smith CC, Dixon RA, Wynne AM, Theodorou L, Ong SG, Subrayan S, Davidson SM, Hausenloy DJ, Yellon DM. Leptin-induced cardioprotection involves JAK/STAT signaling that may be linked to the mitochondrial permeability transition pore. *Am J Physiol Heart Circ Physiol*. 2010;299:H1265–H1270. doi: 10.1152/ajpheart.00092.2010
72. Eguchi M, Liu Y, Shin EJ, Sweeney G. Leptin protects H9c2 rat cardiomyocytes from H2O2-induced apoptosis. *FEBS J*. 2008;275:3136–3144. doi: 10.1111/j.1742-4658.2008.06465.x
73. Lee Y, Naseem RH, Duplomb L, Park BH, Garry DJ, Richardson JA, Schaffer JE, Unger RH. Hyperleptinemia prevents lipotoxic cardiomyopathy in acyl CoA synthase transgenic mice. *Proc Natl Acad Sci USA*. 2004;101:13624–13629. doi: 10.1073/pnas.0405499101
74. Smith CC, Mocanu MM, Davidson SM, Wynne AM, Simpkin JC, Yellon DM. Leptin, the obesity-associated hormone, exhibits direct cardioprotective effects. *Br J Pharmacol*. 2006;149:5–13. doi: 10.1038/sj.bjp.0706834
75. Dong F, Zhang X, Yang X, Esberg LB, Yang H, Zhang Z, Culver B, Ren J. Impaired cardiac contractile function in ventricular myocytes from leptin-deficient ob/ob obese mice. *J Endocrinol*. 2006;188:25–36. doi: 10.1677/joe.1.06241
76. Dong F, Zhang X, Ren J. Leptin regulates cardiomyocyte contractile function through endothelin-1 receptor-NADPH oxidase pathway. *Hypertension*. 2006;47:222–229. doi: 10.1161/01.HYP.0000198555.51645.f1
77. Wold LE, Relling DP, Duan J, Norby FL, Ren J. Abrogated leptin-induced cardiac contractile response in ventricular myocytes under spontaneous hypertension: role of Jak/STAT pathway. *Hypertension*. 2002;39:69–74. doi: 10.1161/hy0102.100777
78. Hall ME, Harmancey R, Stec DE. Lean heart: role of leptin in cardiac hypertrophy and metabolism. *World J Cardiol*. 2015;7:511–524. doi: 10.4330/wjcv.7.511
79. Pieterse C, Schutte R, Schutte AE. Leptin relates to prolonged cardiovascular recovery after acute stress in Africans: The SABPA study. *Nutr Metab Cardiovasc Dis*. 2016;26:45–52. doi: 10.1016/j.numecd.2015.10.014
80. Puurunen VP, Kiviniemi A, Lepojärvi S, Piira OP, Hedberg P, Junttila J, Ukkola O, Huikuri H. Leptin predicts short-term major adverse cardiac events in patients with coronary artery disease. *Ann Med*. 2017;49:448–454. doi: 10.1080/07853890.2017.1301678
81. Tsai JP, Wang JH, Chen ML, Yang CF, Chen YC, Hsu BG. Association of serum leptin levels with central arterial stiffness in coronary artery disease patients. *BMC Cardiovasc Disord*. 2016;16:80. doi: 10.1186/s12872-016-0268-5
82. Singhal A, Farooqi IS, Cole TJ, O'Rahilly S, Fewtrell M, Kattenhorn M, Lucas A, Deanfield J. Influence of leptin on arterial distensibility: a novel link between obesity and cardiovascular disease? *Circulation*. 2002;106:1919–1924. doi: 10.1161/01.cir.0000033219.24717.52
83. Jamar G, Caranti DA, de Cassia Cesar H, Masquio DCL, Bandoni DH, Pisani LP. Leptin as a cardiovascular risk marker in metabolically healthy obese: Hyperleptinemia in metabolically healthy obese. *Appetite*. 2017;108:477–482. doi: 10.1016/j.appet.2016.11.013
84. Aizawa-Abe M, Ogawa Y, Masuzaki H, Ebihara K, Satoh N, Iwai H, Matsuoka N, Hayashi T, Hosoda K, Inoue G, et al. Pathophysiological role of leptin in obesity-related hypertension. *J Clin Invest*. 2000;105:1243–1252. doi: 10.1172/JCI8341
85. Kain D, Simon AJ, Greenberg A, Ben Zvi D, Gilburd B, Schneiderman J. Cardiac leptin overexpression in the context of acute MI and reperfusion potentiates myocardial remodeling and left ventricular dysfunction. *PLoS One*. 2018;13:e0203902. doi: 10.1371/journal.pone.0203902
86. Ben-Zvi D, Savion N, Kolodgie F, Simon A, Fisch S, Schafer K, Bachner-Hinzenon N, Cao X, Gertler A, Solomon G, et al. Local application of leptin antagonist attenuates angiotensin ii-induced ascending aortic aneurysm and cardiac remodeling. *J Am Heart Assoc*. 2016;5:e003474.
87. Purdham DM, Rajapurohitam V, Zeidan A, Huang C, Gross GJ, Karmazyn M. A neutralizing leptin receptor antibody mitigates hypertrophy and hemodynamic dysfunction in the postinfarcted rat heart. *Am J Physiol Heart Circ Physiol*. 2008;295:H441–H446. doi: 10.1152/ajpheart.91537.2007
88. Feijóo-Bandín S, Portolés M, Roselló-Lletí E, Rivera M, González-Juanatey JR, Lago F. 20 years of leptin: role of leptin in cardiomyocyte physiology and pathophysiology. *Life Sci*. 2015;140:10–18. doi: 10.1016/j.lfs.2015.02.016
89. Zhao S, Li N, Zhu Y, Straub L, Zhang Z, Wang MY, Zhu Q, Kusminski CM, Elmquist JK, Scherer PE. Partial leptin deficiency confers resistance to diet-induced obesity in mice. *Mol Metab*. 2020;37:100995. doi: 10.1016/j.molmet.2020.100995
90. Zhao S, Kusminski CM, Elmquist JK, Scherer PE. Leptin: less is more. *Diabetes*. 2020;69:823–829. doi: 10.2337/dbi19-0018
91. Zhao S, Zhu Y, Schultz RD, Li N, He Z, Zhang Z, Caron A, Zhu Q, Sun K, Xiong W, et al. Partial leptin reduction as an insulin sensitization and weight loss strategy. *Cell Metab*. 2019;30:706.e6–719.e6. doi: 10.1016/j.cmet.2019.08.005
92. Knight ZA, Hannan KS, Greenberg ML, Friedman JM. Hyperleptinemia is required for the development of leptin resistance. *PLoS One*. 2010;5:e11376. doi: 10.1371/journal.pone.0011376
93. Berg AH, Combs TP, Du X, Brownlee M, Scherer PE. The adipocyte-secreted protein Acrp30 enhances hepatic insulin action. *Nat Med*. 2001;7:947–953. doi: 10.1038/90992
94. Ye R, Holland WL, Gordillo R, Wang M, Wang QA, Shao M, Morley TS, Gupta RK, Stahl A, Scherer PE. Adiponectin is essential for lipid homeostasis and survival under insulin deficiency and promotes beta-cell regeneration. *Elife*. 2014;3:e03851.
95. Qi Y, Takahashi N, Hileman SM, Patel HR, Berg AH, Pajvani UB, Scherer PE, Ahima RS. Adiponectin acts in the brain to decrease body weight. *Nat Med*. 2004;10:524–529. doi: 10.1038/nm1029
96. Oshima K, Nampei A, Matsuda M, Iwaki M, Fukuhara A, Hashimoto J, Yoshikawa H, Shimomura I. Adiponectin increases bone mass by suppressing osteoclast and activating osteoblast. *Biochem Biophys Res Commun*. 2005;331:520–526. doi: 10.1016/j.bbrc.2005.03.210
97. Zhu Q, Scherer PE. Immunologic and endocrine functions of adipose tissue: implications for kidney disease. *Nat Rev Nephrol*. 2018;14:105–120. doi: 10.1038/nrneph.2017.157

98. Ouchi N, Kobayashi H, Kihara S, Kumada M, Sato K, Inoue T, Funahashi T, Walsh K. Adiponectin stimulates angiogenesis by promoting cross-talk between AMP-activated protein kinase and Akt signaling in endothelial cells. *J Biol Chem*. 2004;279:1304–1309. doi: 10.1074/jbc.M310389200
99. Wolf AM, Wolf D, Rumpold H, Enrich B, Tilg H. Adiponectin induces the anti-inflammatory cytokines IL-10 and IL-1RA in human leukocytes. *Biochem Biophys Res Commun*. 2004;323:630–635. doi: 10.1016/j.bbrc.2004.08.145
100. Pajvani UB, Du X, Combs TP, Berg AH, Rajala MW, Schulthess T, Engel J, Brownlee M, Scherer PE. Structure-function studies of the adipocyte-secreted hormone Acrp30/adiponectin. Implications for metabolic regulation and bioactivity. *J Biol Chem*. 2003;278:9073–9085. doi: 10.1074/jbc.M207198200
101. Silha JV, Krsek M, Skrha JV, Sucharda P, Nyomba BL, Murphy LJ. Plasma resistin, adiponectin and leptin levels in lean and obese subjects: correlations with insulin resistance. *Eur J Endocrinol*. 2003;149:331–335. doi: 10.1530/eje.0.1490331
102. Doumatey AP, Bentley AR, Zhou J, Huang H, Adeyemo A, Rotimi CN. Paradoxical hyperadiponectinemia is associated with the Metabolically Healthy Obese (MHO) phenotype in african americans. *J Endocrinol Metab*. 2012;2:51–65. doi: 10.4021/jem95W
103. Marangoni RG, Masui Y, Fang F, Korman B, Lord G, Lee J, Lakota K, Wei J, Scherer PE, Otvos L, et al. Adiponectin is an endogenous antifibrotic mediator and therapeutic target. *Sci Rep*. 2017;7:4397. doi: 10.1038/s41598-017-04162-1
104. Straub LG, Scherer PE. Metabolic messengers: adiponectin. *Nat Metab*. 2019;1:334–339. doi: 10.1038/s42255-019-0041-z
105. Maeda N, Shimomura I, Kishida K, Nishizawa H, Matsuda M, Nagaretani H, Furuyama N, Kondo H, Takahashi M, Arita Y, et al. Diet-induced insulin resistance in mice lacking adiponectin/ACRP30. *Nat Med*. 2002;8:731–737. doi: 10.1038/nm724
106. Kubota N, Terauchi Y, Yamauchi T, Kubota T, Moroi M, Matsui J, Eto K, Yamashita T, Kamon J, Satoh H, et al. Disruption of adiponectin causes insulin resistance and neointimal formation. *J Biol Chem*. 2002;277:25863–25866. doi: 10.1074/jbc.C200251200
107. Xia JY, Sun K, Hepler C, Ghaben AL, Gupta RK, An YA, Holland WL, Morley TS, Adams AC, Gordillo R, et al. Acute loss of adipose tissue-derived adiponectin triggers immediate metabolic deterioration in mice. *Diabetologia*. 2018;61:932–941. doi: 10.1007/s00125-017-4516-8
108. Combs TP, Berg AH, Obici S, Scherer PE, Rossetti L. Endogenous glucose production is inhibited by the adipose-derived protein Acrp30. *J Clin Invest*. 2001;108:1875–1881. doi: 10.1172/JCI14120
109. Yamauchi T, Nio Y, Maki T, Kobayashi M, Takazawa T, Iwabu M, Okada-Iwabu M, Kawamoto S, Kubota N, Kubota T, et al. Targeted disruption of AdipoR1 and AdipoR2 causes abrogation of adiponectin binding and metabolic actions. *Nat Med*. 2007;13:332–339. doi: 10.1038/nm1557
110. Holland WL, Xia JY, Johnson JA, Sun K, Pearson MJ, Sharma AX, Quittner-Strom E, Tippetts TS, Gordillo R, Scherer PE. Inducible overexpression of adiponectin receptors highlight the roles of adiponectin-induced ceramidase signaling in lipid and glucose homeostasis. *Mol Metab*. 2017;6:267–275. doi: 10.1016/j.molmet.2017.01.002
111. Okada-Iwabu M, Yamauchi T, Iwabu M, Honma T, Hamagami K, Matsuda K, Yamaguchi M, Tanabe H, Kimura-Someya T, Shirouzu M, et al. A small-molecule AdipoR agonist for type 2 diabetes and short life in obesity. *Nature*. 2013;503:493–499. doi: 10.1038/nature12656
112. Lee CH, Lui DTW, Cheung CYY, Fong CHY, Yuen MMA, Chow WS, Woo YC, Xu A, Lam KSL. Higher circulating adiponectin concentrations predict incident cancer in type 2 diabetes - the adiponectin paradox. *J Clin Endocrinol Metab*. 2020;105:dga075.
113. Baker JF, Newman AB, Kanaya A, Leonard MB, Zemel B, Miljkovic I, Long J, Weber D, Harris TB. The adiponectin paradox in the elderly: associations with body composition, physical functioning, and mortality. *J Gerontol A Biol Sci Med Sci*. 2019;74:247–253. doi: 10.1093/gerona/gly017
114. Kobayashi H, Ouchi N, Kihara S, Walsh K, Kumada M, Abe Y, Funahashi T, Matsuzawa Y. Selective suppression of endothelial cell apoptosis by the high molecular weight form of adiponectin. *Circ Res*. 2004;94:e27–e31. doi: 10.1161/01.RES.0000119921.86460.37
115. Natarajan R, Salloum FN, Fisher BJ, Kukreja RC, Fowler AA III. Hypoxia inducible factor-1 upregulates adiponectin in diabetic mouse hearts and attenuates post-ischemic injury. *J Cardiovasc Pharmacol*. 2008;51:178–187. doi: 10.1097/FJC.0b013e31815f248d
116. Khan RS, Kato TS, Chokshi A, Chew M, Yu S, Wu C, Singh P, Cheema FH, Takayama H, Harris C, et al. Adipose tissue inflammation and adiponectin resistance in patients with advanced heart failure: correction after ventricular assist device implantation. *Circ Heart Fail*. 2012;5:340–348. doi: 10.1161/CIRCHEARTFAILURE.111.964031
117. Halberg N, Schraw TD, Wang ZV, Kim JY, Yi J, Hamilton MP, Luby-Phelps K, Scherer PE. Systemic fate of the adipocyte-derived factor adiponectin. *Diabetes*. 2009;58:1961–1970. doi: 10.2337/db08-1750
118. Tacke F, Wüstefeld T, Horn R, Luedde T, Srinivas Rao A, Manns MP, Trautwein C, Brabant G. High adiponectin in chronic liver disease and cholestasis suggests biliary route of adiponectin excretion in vivo. *J Hepatol*. 2005;42:666–673. doi: 10.1016/j.jhep.2004.12.024
119. Kizer JR, Arnold AM, Strotmeyer ES, Ives DG, Cushman M, Ding J, Kritchevsky SB, Chaves PH, Hirsch CH, Newman AB. Change in circulating adiponectin in advanced old age: determinants and impact on physical function and mortality. The Cardiovascular Health Study All Stars Study. *J Gerontol A Biol Sci Med Sci*. 2010;65:1208–1214. doi: 10.1093/gerona/glq122
120. Combs TP, Wagner JA, Berger J, Doebber T, Wang WJ, Zhang BB, Tanen M, Berg AH, O'Rahilly S, Savage DB, et al. Induction of adipocyte complement-related protein of 30 kilodaltons by PPARγ agonists: a potential mechanism of insulin sensitization. *Endocrinology*. 2002;143:998–1007. doi: 10.1210/endo.143.3.8662
121. Tsutomoto T, Tanaka T, Sakai H, Ishikawa C, Fujii M, Yamamoto T, Horie M. Total and high molecular weight adiponectin, haemodynamics, and mortality in patients with chronic heart failure. *Eur Heart J*. 2007;28:1723–1730. doi: 10.1093/eurheartj/ehm154
122. Karas MG, Benkeser D, Arnold AM, Bartz TM, Djousse L, Mukamal KJ, Ix JH, Ziemann SJ, Siscovick DS, Tracy RP, et al. Relations of plasma total and high-molecular-weight adiponectin to new-onset heart failure in adults ≥65 years of age (from the Cardiovascular Health study). *Am J Cardiol*. 2014;113:328–334. doi: 10.1016/j.amjcard.2013.09.027
123. Lin H, Lian WS, Chen HH, Lai PF, Cheng CF. Adiponectin ameliorates iron-overload cardiomyopathy through the PPARα-PGC-1-dependent signaling pathway. *Mol Pharmacol*. 2013;84:275–285. doi: 10.1124/mol.112.083964
124. Nelson MD, Victor RG, Szczepaniak EW, Simha V, Garg A, Szczepaniak LS. Cardiac steatosis and left ventricular hypertrophy in patients with generalized lipodystrophy as determined by magnetic resonance spectroscopy and imaging. *Am J Cardiol*. 2013;112:1019–1024. doi: 10.1016/j.amjcard.2013.05.036
125. Bhayana S, Siu VM, Joubert GI, Clarson CL, Cao H, Hegele RA. Cardiomyopathy in congenital complete lipodystrophy. *Clin Genet*. 2002;61:283–287. doi: 10.1034/j.1399-0004.2002.610407.x
126. Mazumder PK, O'Neill BT, Roberts MW, Buchanan J, Yun UJ, Cooksey RC, Boudina S, Abel ED. Impaired cardiac efficiency and increased fatty acid oxidation in insulin-resistant ob/ob mouse hearts. *Diabetes*. 2004;53:2366–2374. doi: 10.2337/diabetes.53.9.2366
127. Oral EA, Simha V, Ruiz E, Andewelt A, Premkumar A, Snell P, Wagner AJ, DePaoli AM, Reitman ML, Taylor SI, et al. Leptin-replacement therapy for lipodystrophy. *N Engl J Med*. 2002;346:570–578. doi: 10.1056/NEJMoa012437
128. Shinmura K, Tamaki K, Sano M, Murata M, Yamakawa H, Ishida H, Fukuda K. Impact of long-term caloric restriction on cardiac senescence: caloric restriction ameliorates cardiac diastolic dysfunction associated with aging. *J Mol Cell Cardiol*. 2011;50:117–127. doi: 10.1016/j.jmcc.2010.10.018
129. Han X, Turdi S, Hu N, Guo R, Zhang Y, Ren J. Influence of long-term caloric restriction on myocardial and cardiomyocyte contractile function and autophagy in mice. *J Nutr Biochem*. 2012;23:1592–1599. doi: 10.1016/j.jnutbio.2011.11.002
130. Robertson LT, Treviño-Villarreal JH, Mejia P, Grondin Y, Harputlugil E, Hine C, Vargas D, Zheng H, Ozaki CK, Kristal BS, et al. Protein and calorie restriction contribute additively to protection from renal ischemia reperfusion injury partly via leptin reduction in male mice. *J Nutr*. 2015;145:1717–1727. doi: 10.3945/jn.114.199380
131. Rogozina OP, Bonorden MJ, Seppanen CN, Grande JP, Cleary MP. Effect of chronic and intermittent calorie restriction on serum adiponectin and leptin and mammary tumorigenesis. *Cancer Prev Res (Phila)*. 2011;4:568–581. doi: 10.1158/1940-6207.CAPR-10-0140
132. Racil G, Coquart JB, Elmontassar W, Haddad M, Goebel R, Chauouchi A, Amri M, Chamari K. Greater effects of high- compared with moderate-intensity interval training on cardio-metabolic variables, blood leptin concentration and ratings of perceived exertion in obese adolescent females. *Biol Sport*. 2016;33:145–152. doi: 10.5604/20831862.1198633
133. Verma S, Poulter NR, Bhatt DL, Bain SC, Buse JB, Leiter LA, Nauck MA, Pratley RE, Zinman B, Ørsted DD, et al. Effects of liraglutide on cardiovascular outcomes in patients with type 2 diabetes mellitus with or without

- history of myocardial infarction or stroke. *Circulation*. 2018;138:2884–2894. doi: 10.1161/CIRCULATIONAHA.118.034516
134. Wu P, Wen W, Li J, Xu J, Zhao M, Chen H, Sun J. Systematic review and meta-analysis of randomized controlled trials on the effect of SGLT2 inhibitor on blood leptin and adiponectin level in patients with type 2 diabetes. *Horm Metab Res*. 2019;51:487–494. doi: 10.1055/a-0958-2441
 135. Packer M. Do sodium-glucose co-transporter-2 inhibitors prevent heart failure with a preserved ejection fraction by counterbalancing the effects of leptin? A novel hypothesis. *Diabetes Obes Metab*. 2018;20:1361–1366. doi: 10.1111/dom.13229
 136. Matthews VB, Elliot RH, Rudnicka C, Hricova J, Herat L, Schlaich MP. Role of the sympathetic nervous system in regulation of the sodium glucose cotransporter 2. *J Hypertens*. 2017;35:2059–2068. doi: 10.1097/HJH.0000000000001434
 137. Krysiak R, Sierant M, Marek B, Bienek R, Okopień B. The effect of angiotensin-converting enzyme inhibitors on plasma adipokine levels in normotensive patients with coronary artery disease. *Endokrynol Pol*. 2010;61:280–287.
 138. Singh P, Zhang Y, Sharma P, Covassin N, Soucek F, Friedman PA, Somers VK. Statins decrease leptin expression in human white adipocytes. *Physiol Rep*. 2018;6:e13566.
 139. Mick GJ, Wang X, Ling Fu C, McCormick KL. Inhibition of leptin secretion by insulin and metformin in cultured rat adipose tissue. *Biochim Biophys Acta*. 2000;1502:426–432. doi: 10.1016/s0925-4439(00)00074-0
 140. Lazzari P, Sanna A, Mastinu A, Cabasino S, Manca I, Pani L. Weight loss induced by rimonabant is associated with an altered leptin expression and hypothalamic leptin signaling in diet-induced obese mice. *Behav Brain Res*. 2011;217:432–438. doi: 10.1016/j.bbr.2010.11.022
 141. Tam J, Cinar R, Liu J, Godlewski G, Wesley D, Jourdan T, Szanda G, Mukhopadhyay B, Chedester L, Liow JS, et al. Peripheral cannabinoid-1 receptor inverse agonism reduces obesity by reversing leptin resistance. *Cell Metab*. 2012;16:167–179. doi: 10.1016/j.cmet.2012.07.002
 142. Yang WS, Jeng CY, Wu TJ, Tanaka S, Funahashi T, Matsuzawa Y, Wang JP, Chen CL, Tai TY, Chuang LM. Synthetic peroxisome proliferator-activated receptor-gamma agonist, rosiglitazone, increases plasma levels of adiponectin in type 2 diabetic patients. *Diabetes Care*. 2002;25:376–380. doi: 10.2337/diacare.25.2.376
 143. Zhang N, Wei WY, Liao HH, Yang Z, Hu C, Wang SS, Deng W, Tang QZ. AdipoRon, an adiponectin receptor agonist, attenuates cardiac remodeling induced by pressure overload. *J Mol Med (Berl)*. 2018;96:1345–1357. doi: 10.1007/s00109-018-1696-8
 144. Hug C, Wang J, Ahmad NS, Bogan JS, Tsao TS, Lodish HF. T-cadherin is a receptor for hexameric and high-molecular-weight forms of Acrp30/adiponectin. *Proc Natl Acad Sci USA*. 2004;101:10308–10313. doi: 10.1073/pnas.0403382101
 145. Denzel MS, Scimia MC, Zumstein PM, Walsh K, Ruiz-Lozano P, Ranscht B. T-cadherin is critical for adiponectin-mediated cardioprotection in mice. *J Clin Invest*. 2010;120:4342–4352. doi: 10.1172/JCI43464
 146. Selthofer-Relatić K, Radić R, Stupin A, Šišljagić V, Bošnjak I, Bulj N, Selthofer R, Delić Brkljačić D. Leptin/adiponectin ratio in overweight patients – gender differences. *Diab Vasc Dis Res*. 2018;15:260–262. doi: 10.1177/1479164117752491
 147. Finucane FM, Luan J, Wareham NJ, Sharp SJ, O'Rahilly S, Balkau B, Flyvbjerg A, Walker M, Højlund K, Nolan JJ, et al; (on behalf of the European Group for the Study of Insulin Resistance: Relationship between Insulin Sensitivity and Cardiovascular Disease Risk Study Group). Correlation of the leptin:adiponectin ratio with measures of insulin resistance in non-diabetic individuals. *Diabetologia*. 2009;52:2345–2349. doi: 10.1007/s00125-009-1508-3
 148. Norata GD, Raselli S, Grigore L, Garlaschelli K, Dozio E, Magni P, Catapano AL. Leptin:adiponectin ratio is an independent predictor of intima media thickness of the common carotid artery. *Stroke*. 2007;38:2844–2846. doi: 10.1161/STROKEAHA.107.485540
 149. López-Jaramillo P, Gómez-Arbeláez D, López-López J, López-López C, Martínez-Ortega J, Gómez-Rodríguez A, Triana-Cubillos S. The role of leptin/adiponectin ratio in metabolic syndrome and diabetes. *Horm Mol Biol Clin Invest*. 2014;18:37–45. doi: 10.1515/hmbci-2013-0053
 150. Saltiel AR, Olefsky JM. Inflammatory mechanisms linking obesity and metabolic disease. *J Clin Invest*. 2017;127:1–4. doi: 10.1172/JCI92035
 151. Zaletel J, Barlovic DP, Prezelj J. Adiponectin-leptin ratio: a useful estimate of insulin resistance in patients with Type 2 diabetes. *J Endocrinol Invest*. 2010;33:514–518. doi: 10.1007/BF03346639
 152. Magkos F, Fraterrigo G, Yoshino J, Luecking C, Kirbach K, Kelly SC, de Las Fuentes L, He S, Okunade AL, Patterson BW, et al. Effects of moderate and subsequent progressive weight loss on metabolic function and adipose tissue biology in humans with obesity. *Cell Metab*. 2016;23:591–601. doi: 10.1016/j.cmet.2016.02.005
 153. Perego L, Pizzocri P, Corradi D, Maisano F, Paganelli M, Fiorina P, Barbieri M, Morabito A, Paolisso G, Folli F, et al. Circulating leptin correlates with left ventricular mass in morbid (grade III) obesity before and after weight loss induced by bariatric surgery: a potential role for leptin in mediating human left ventricular hypertrophy. *J Clin Endocrinol Metab*. 2005;90:4087–4093. doi: 10.1210/jc.2004-1963
 154. Faraj M, Havel RJ, Phélis S, Blank D, Sniderman AD, Cianflone K. Plasma acylation-stimulating protein, adiponectin, leptin, and ghrelin before and after weight loss induced by gastric bypass surgery in morbidly obese subjects. *J Clin Endocrinol Metab*. 2003;88:1594–1602. doi: 10.1210/jc.2002-021309
 155. Jin T, Song Z, Weng J, Fantus IG. Curcumin and other dietary polyphenols: potential mechanisms of metabolic actions and therapy for diabetes and obesity. *Am J Physiol Endocrinol Metab*. 2018;314:E201–E205. doi: 10.1152/ajpendo.00285.2017
 156. Drucker DJ. Advances in oral peptide therapeutics. *Nat Rev Drug Discov*. 2020;19:277–289. doi: 10.1038/s41573-019-0053-0
 157. Wolfrum C, Straub LG. Lessons from cre-mice and indicator mice. *Handb Exp Pharmacol*. 2019;251:37–54. doi: 10.1007/164_2018_146